# Crotoxin, the major toxin from the rattlesnake *Crotalus durissus terrificus*, inhibits <sup>3</sup>H-choline uptake in guinea pig ileum

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#### **Abstract**

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We examined the effect of crotoxin, the neurotoxic complex from the venom of the South American rattlesnake Crotalus durissus terrificus, on the uptake of <sup>3</sup>H-choline in minces of smooth muscle myenteric plexus from guinea pig ileum. In the concentration range used (0.03-1 μM) and up to 10 min of treatment, crotoxin decreased <sup>3</sup>H-choline uptake by 50-75% compared to control. This inhibition was time dependent and did not seem to be associated with the disruption of the neuronal membrane, because at least for the first 20 min of tissue exposure to the toxin (up to 1 µM) the levels of lactate dehydrogenase (LDH) released into the supernatant were similar to those of controls. Higher concentrations of crotoxin or more extensive incubation times with this toxin resulted in elevation of LDH activity detected in the assay supernatant. The inhibitory effect of crotoxin on <sup>3</sup>H-choline uptake seems to be associated with its phospholipase activity since the equimolar substitution of Sr<sup>2+</sup> for Ca<sup>2+</sup> in the incubation medium or the modification of the toxin with p-bromophenacyl bromide substantially decreased this effect. Our results show that crotoxin inhibits <sup>3</sup>Hcholine uptake with high affinity (EC<sub>25</sub> =  $10 \pm 5$  nM). We suggest that this inhibition could explain, at least in part, the blocking effect of crotoxin on neurotransmission.

#### **Key words**

- Crotoxin
- · Choline uptake
- Guinea pig ileum
- Crotalus durissus terrificus
- Phospholipase A<sub>2</sub>

Crotoxin from the venom of the South American rattlesnake *Crotalus durissus ter- rificus* (1) is a complex of two different subunits: a basic and weakly toxic phospholipase A<sub>2</sub>, component B (CB), and an acidic, non-toxic protein, component A (CA) (2). Crotoxin exerts its pathophysiological action by blocking neuromuscular transmission. It acts primarily at the presynaptic level by altering neurotransmitter release (3,4) al-

though it is also able to act postsynaptically by stabilizing the acetylcholine receptor in an inactive state (5). Crotoxin blocks acetylcholine release induced by *Tityus serrulatus* venom in guinea pig ileum (6) and the response of smooth muscle myenteric plexus to field electrical stimulation (7,8).

In the present study we determined the effect of crotoxin on the transport of choline by innervated longitudinal muscle minces of 1094 L.S. Kattah et al.

guinea pig small intestine. We found that crotoxin partially inhibited <sup>3</sup>H-choline uptake and propose that this may be related to the neuromuscular blockade caused by the toxin. Parts of the present data have appeared previously in different form (9).

The <sup>3</sup>H-choline uptake assays were carried out by the method of Pert and Snyder (10), with minor modifications. Briefly, fragments of smooth muscle myenteric plexus were weighed and minced with scissors into small pieces, and samples (about 40 mg of tissue) in triplicate were suspended in 1.0 ml depolarizing buffer (Tyrode solution containing 50 mM KCl, with equimolar reduction of NaCl), at 37°C for 15 min upon agitation, to release endogenous acetylcholine. After washing with 1 ml Tyrode solution (composition: 136 mM NaCl, 2.7 mM KCl, 1.8 mM CaCl<sub>2</sub>, 5.5 mM glucose, 10 mM Tris, and 20 µM paraoxan, pH 7.4), minces were incubated for 10 min in 1 ml of normal buffer at 37°C, with or without crotoxin. One µCi <sup>3</sup>H-choline (12 nM) in 1.0 ml of normal or depolarizing incubation buffer was added to the sample and the incubation prolonged for 10 min more. Incubations were stopped by the rapid addition of icecold Tyrode solution with 1.0 µM unlabeled choline and by centrifugation (13,000 rpm, 5 min) at 4°C using a Marathon 13K/H microcentrifuge (Marathon, Pittsburg, PA, USA). Nonspecific or passive uptake was defined as uptake occurring at 4°C. The <sup>3</sup>H-choline uptake is expressed as cpm or as percentage of control at 37°C. The tissue was solubilized with 50 µl 1 M NaOH for 40 min at 60°C. After dilution with 1 ml Tyrode and homogenization, 250-µl samples were counted by liquid scintillation spectrometry in a liquid scintillation counter (LKB/Walack, Bromma, Sweden).

All experiments were performed on preparations obtained from male and female guinea pigs (250-600 g). The animals were anesthetized with CO<sub>2</sub>, killed by a blow to the head and bled. The entire small intestine was

rapidly removed and washed with Tyrode solution at 37°C. The longitudinal muscle strips containing adherent (Auerbach's) myenteric plexus were obtained by the method of Paton and Zar (11).

In some experiments, the supernatants were used for the determination of lactate dehydrogenase (LDH) activity according to Kubowitz and Ott (12). Briefly, an aliquot from the supernatant (150 µl) was placed in a cuvette containing LDH assay buffer (50 mM phosphate, 0.6 mM sodium pyruvate and 0.2 mM NADH). The absorbance (365 nm) was recorded at 1, 2 and 3 min using a Shimadzu UV-160A spectrophotometer and the enzymatic activity was calculated. One unit of enzymatic activity is defined as that producing oxidation of 1 µM NADH/min at 25°C. Total lysis, corresponding to 100% LDH in minces, was determined after treatment with 1% Triton X-100.

The alkylation by p-bromophenacyl bromide was performed at 30°C in 1.5 ml of 100 mM sodium cacodylate buffer, pH 5.8, and 100 mM NaCl. The phospholipase isolated from the crotoxin complex (15 µM) was preincubated for 1 h under these conditions before the reaction was started by adding 15 µl of 8 mM p-bromophenacyl bromide in acetone. After incubation at 30°C for 60 min, excess reagent was eliminated by dialysis against water. The crotoxin complex was reconstituted by adding CA (in 1.5 ml 50 mM Tris-HCl, pH 7.4, 0.1 M NaCl) to CB (1.5 CA:1.0 CB) treated with p-bromophenacyl bromide. All assays were done in triplicate and repeated at least three times (N = 3).

Data were analyzed statistically by the Tukey test, Dunnett test, Student *t*-test, and F test (13). The test performed for each experiment is described in the legends to the figures.

<sup>3</sup>H-Choline uptake and the release of LDH, a cytoplasmic enzyme marker for tissue integrity, were monitored at different time intervals (Figure 1A and B). The levels of LDH measured in samples treated for 20

min with 0.3 µM crotoxin were similar to those of the control and amounted to about 20% (Figure 1B), while the inhibitory effect on <sup>3</sup>H-choline uptake under the same conditions was about 50%. After a longer incubation time (e.g., 120 min) with 0.3 µM crotoxin (the concentration used in most experiments), <sup>3</sup>H-choline uptake was drastically reduced to values similar to those measured at 4°C (Figure 1A, inset), while LDH activity increased to 40% in the supernatant. Crotoxin (0.3 μM) inhibited <sup>3</sup>H-choline uptake after 2-4 min of incubation, whereas the increase in LDH activity was detected several minutes later (data not shown). The calculated EC<sub>25</sub>  $(10 \pm 5 \text{ nM})$  showed a high affinity of crotoxin for this preparation (data not shown). Different controls (inset in Figure 1) were regularly performed in all experiments and included incubations at 4°C and at 37°C in normal and in depolarizing medium. About 20-30% uptake was observed in normal medium at 4°C. These values were interpreted as passive uptake. In depolarizing medium the choline transporter was inhibited as described (14). The difference between total uptake measured at 37°C and passive uptake measured at 4°C corresponds to high affinity choline transport. This type of transport is characteristic of nerve cells (10). The total uptake at 37°C was considered as 100%.

To determine if the phospholipase activity from crotoxin was implicated in the inhibition of <sup>3</sup>H-choline uptake, two different experiments were carried out. The modification of crotoxin with p-bromophenacyl bromide, which inactivates the phospholipase A<sub>2</sub> activity of crotoxin (15), decreased the inhibitory effect of the toxin from 50% to about 20% (Figure 2A). The equimolar substitution of Sr<sup>2+</sup>, a competitive inhibitor of Ca<sup>2+</sup> for phospholipase activity, for Ca<sup>2+</sup> with the addition of 1 mM EGTA in the medium also decreased this effect of the toxin (Figure 2B). Note that in both conditions crotoxin still showed a partial, but not significant inhibitory effect. The existence

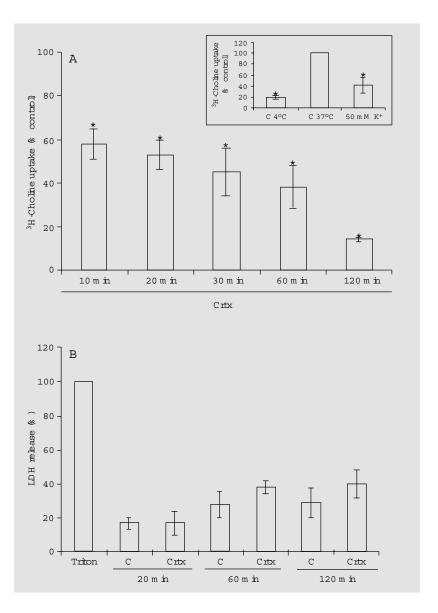
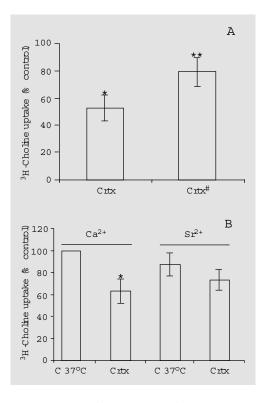


Figure 1 - Effect of pre-incubation time with crotoxin (Crtx) on <sup>3</sup>H -choline uptake inhibition and lactate dehydrogenase (LDH) release. A, M inces of neurom uscular preparation (40 m g in triplicate) were pre-incubated for different periods of time at  $37^{\circ}\text{C}$  in Tyrode solution in the presence of  $0.3 \, \mu M$  Crtx.  $^3H$  -Choline (1.2 nM) was added and the uptake measured after 10 m in incubation. Values are reported as percentage of total uptake (control at 37°C) and represent the mean and standard deviation of the mean of 3 different experiments. \*P<0.05 com pared to control (uptake at  $37^{\circ}$ C). Analysis of variance (P = 0) show ed that there w as a difference between treatments. Using the F test for repeated measures and calculating the reverse Helm et contrasts we observed that: control>10' (P = 0.012), 10' > 20' (P = 0.014), 20' > 30' (P = 0.009), 30' > 60' (P = 0.024) and 60' > 120' (P = 0.006).Inset: Control (C) values obtained in 3 different experiments at 37°C (100% uptake) in norm aland depolarizing medium (50 m M K+) and at 4°C in norm alm edium, for 10 m in after the addition of  $^3H$  -choline (12 nM ). \*P < 0.05 com pared to control at  $37^{\circ}C$  (t-test). B, LDH released in the control condition (C) and upon incubation with Crtx (0.3 µM) for different periods of time at 37°C. The LDH released by the preparation treated with 1% Triton X-100 was taken as 100%. Values (mean and standard deviation of the mean of 3 different experin ents) were not significant compared to the respective controls (P>0.05).

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of a residual phospholipase activity of crotoxin under our experimental conditions could explain this result. The inhibitory effect of crotoxin on the release of acetylcholine induced by tityustoxin in this same neuromuscular preparation has also been suggested to depend on its phospholipase activity (6). The inhibitory effect of some fatty acids on choline uptake has already been shown in rat brain synaptosomes (16) and was confirmed in preliminary experiments using our preparation (data not shown). This effect also argues in favor of a role of the phospholipase activity of crotoxin on the inhibition process. The fatty acids released

Figure 2 - Inhibition of the phospholipase activity of crotoxin Crtx) decreases its effection 3H choline uptake.A, Pre-incubation of minces (40 mg, triplicate) in normal Tyrode solution for 10 m in at 37°C with 1 µM crotoxin m odified with p-brom ophenacyl brom ide (Crtx#) (see M ethods) or with native toxin (Crtx). Values represent the m ean and standard deviation of the m ean of 3 different experim ents. \*P<0.05 com pared to the control at  $37^{\circ}C$ . \*\*P<0.05 compared to the native crotoxin. Analysis of variance showed that there was a difference between treatments (P = 0). M ultiple com parisons based on the confidence intervals for paired differences using the Tukey test (total level of significance = 5% ) showed: control (C)>Crtx; C>Crtx#, and Crtx#> Crtx. B, M inces (40 mg, triplicate) were pre-incubated in norm al Typode solution (with Ca2+) or in modified Typode solution (Sr2+ w as substituted for Ca2+ at



equim olar concentration and 1 m M EGTA w as added), in the absence or in the presence of 0.3  $\mu$ M Crtx, as indicated, for 10 m in at 37°C.  $^3H$  Choline (1.2 nM ) w as added and the uptake m easured after 10 m in incubation. Values represent the m ean and standard deviation of the m ean of 3 different experim ents. \*P<0.05 com pared to the control at 37°C incubated w ith Ca²+ (Studentt-test). by crotoxin phospholipase activity on membrane phospholipids may be implicated in the inhibition of choline uptake.

The role of phospholipase  $A_2$  activity from crotoxin in neurotransmission inhibition at the presynaptic (4,17) and postsynaptic (18) level has been widely supported.

The precise mechanism involving blockade of neurotransmission by crotoxin, responsible for death after the bite of C. durissus terrificus, still is an unsolved question. Studies with β-neurotoxins suggest that the effects observed are related to the phospholipase activity of the toxins or may result from the interactions of toxins with acceptors on biological membranes (19). Toxins that act presynaptically such as \( \beta \- bungarotoxin \), notexin and taipoxin inhibit the release of acetylcholine and are potent inhibitors of the high affinity choline transport system in a membrane preparation derived from the cholinergic endings of the Torpedo electric organ (20). B-Bungarotoxin also inhibits high affinity choline uptake in a synaptosomal preparation of rat cerebral cortices. The presynaptic inhibition of neurotransmitter release seems to be the main effect explaining the neuromuscular blockade induced by crotoxin. In summary, our results indicate that the inhibition of high affinity choline uptake may constitute an important effect of crotoxin, contributing to the neuromuscular blockade it induces.

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